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## 2 **Na<sup>+</sup>/H<sup>+</sup> Exchangers, Structure, 3 and Function: Role in Human Health 4 and Disease**

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## 8 **Synonyms**

[Au2]

9 [Membrane proteins; Na<sup>+</sup>/H<sup>+</sup> exchanger; pH regulation](#)

## 10 **Definition**

11 Na<sup>+</sup>/H<sup>+</sup> exchangers are a family of membrane proteins  
12 that exchange sodium for protons across lipid bilayers.  
13 They are widely distributed in all living cell types and  
14 are critical in several human diseases. The best known  
15 type of Na<sup>+</sup>/H<sup>+</sup> exchanger in multicellular species is  
16 the mammalian Na<sup>+</sup>/H<sup>+</sup> exchanger isoform 1 (NHE1).  
17 It is a plasma membrane protein that regulates intra-  
18 cellular pH by removing one intracellular hydrogen ion  
19 in exchange for one extracellular sodium ion. NHE1  
20 regulates intracellular pH, but is also involved several  
21 diseases in the myocardium and in cancer, in addition  
22 to its role in cell growth, movement and differentia-  
23 tion. This review summarizes current knowledge of  
24 Na<sup>+</sup>/H<sup>+</sup> exchangers, with emphasis on the most well  
25 characterized human Na<sup>+</sup>/H<sup>+</sup> exchanger, the NHE1  
26 isoform.

## Introduction, Na<sup>+</sup>/H<sup>+</sup> Exchangers

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Na<sup>+</sup>/H<sup>+</sup> exchangers are a class of membrane proteins 28  
that exchange Na<sup>+</sup> for H<sup>+</sup>'s across lipid bilayers. The 29  
prokaryotic and eukaryotic genes that encode this 30  
monovalent cation proton antiporter (CPA) superfam- 31  
ily were recently reviewed (Brett et al. 2005). Briefly, 32  
the superfamily includes the CPA1, CPA2, and NaT- 33  
DC (Na<sup>+</sup>-transporting carboxylic acid decarboxylase) 34  
families, each of which has unique bacterial ancestors. 35  
The CPA1 family includes bacterial NhaP transporters. 36  
It also includes many well studied Na<sup>+</sup>/H<sup>+</sup> exchangers 37  
including from fungi, plants and mammals, and 38  
the human NHE1–NHE10 isoforms, the SLC9A 39  
paralogous genes. The CPA2 family shares its origins 40  
with prokaryotic NhaA. The *E. coli* form of NhaA is 41  
structurally, the most well studied Na<sup>+</sup>/H<sup>+</sup> exchanger 42  
and its crystal structure has been deduced. However, 43  
many other genes of this family are not well studied. 44  
This family also includes HsNHA1 and HsNHA2 45  
which are two relatively recently characterized forms 46  
of human Na<sup>+</sup>/H<sup>+</sup> exchanger that may be involved in 47  
hypertension (Brett et al. 2005). The NaT-DC family is 48  
a smaller family that mediates transmembrane export 49  
of 1–2 Na<sup>+</sup> in exchange for an extracellular H<sup>+</sup> 50  
(Brett et al. 2005). 51

## Human NHE1–10

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1. *General* – Human NHE1–NHE10 are membrane 53  
proteins that transport hydrogen ions in exchange 54  
for sodium ions. Human NHE1 is ubiquitously 55  
expressed in mammalian cells and is widespread 56  
throughout the animal kingdom. NHE1 is critical 57

58 in intracellular pH (pH<sub>i</sub>) regulation, protecting cells  
59 from acidification, as well as regulating cell volume  
60 and sodium fluxes (reviewed in). Of the family of  
61 ten isoforms, the NHE1 isoform is ubiquitous and is  
62 on the plasma membrane of mammalian cells.  
63 NHE1 is important in the myocardium as it is impli-  
64 cated in the diseases ischemia/reperfusion injury,  
65 and in heart hypertrophy (see below and reviewed  
66 in Fliegel 2008). NHE1 has a 500 amino acid mem-  
67 brane domain that transports ions and a cytosolic  
68 regulatory domain of approximately 315 amino  
69 acids that modulates activity of the membrane  
70 domain and are a target regulation by proteins and  
71 phosphorylation (Fliegel 2008).

- 72 2. *Subtypes* – Human NHE1 is an 815 amino acid  
73 protein. The family of mammalian NHE-like pro-  
74 teins includes ten isoforms (NHE1–10). Each are  
75 the product of a different gene and with different  
76 tissue distributions and physiological roles. The  
77 first type cloned was named NHE1. NHE2–4 are  
78 principally expressed in the kidney and gastrointes-  
79 tinal tract. NHE5 is located in the brain. NHE6–9  
80 are found in intracellular organelle membranes,  
81 such as endosomes, mitochondria, and the golgi  
82 apparatus, NHE10 is found in osteoclasts. The iden-  
83 tity of the various isoforms varies from 25%  
84 to 70%; however, all have a similar predicted  
85 secondary structure (see below) (Fliegel 2008).

## 86 Structure

- 87 1. *Topological models of NHE1* – Currently, the high-  
88 resolution structure of NHE1 has not yet been  
89 solved, due to the difficulty in expressing and crys-  
90 tallizing membrane proteins. The first 500 residues  
91 are predicted to be 12 transmembrane spanning  
92 segments, and the remaining 315 residues constitute  
93 a cytosolic intracellular regulatory domain (Fliegel  
94 2008). Most of the information on structure of the  
95 membrane domain comes from topology models of  
96 NHE1, some biochemical studies, a low resolution  
97 electron diffraction envelope, a crystal structure of  
98 a bacterial homologue and NMR studies of  
99 fragments of the NHE1 protein. Our laboratory  
100 produced and purified the full length NHE1 protein  
101 (Fliegel 2008). The EM structure showed that  
102 NHE1 exists as a homodimers which was confirmed

103 by intermolecular cross-linking (Fliegel 2008;  
104 Lee et al. 2011).

105 Two initial studies investigated the topology of  
106 the membrane domain (Landau et al. 2007;  
107 Wakabayashi et al. 2000) and vary somewhat  
108 between each other and are reviewed in detail in  
109 (Lee et al. 2011). The topology was first examined  
110 in detail by Wakabayashi et al. (Wakabayashi et al.  
111 2000) and using substituted cysteine accessibility  
112 they suggested that NHE1 contains 12 TM helices  
113 with both the N- and C-terminal in the cytoplasm,  
114 and the N-linked glycosylation site on the extracel-  
115 lular side (Fig. 1). They also suggested that two  
116 intracellular loops and one extracellular loop may  
117 be pore or membrane associated.

118 A more recent model for the topology of NHE1 was  
119 developed by Landau et al. (Landau et al. 2007) and  
120 was based on computational methods, including  
121 evolutionary conservation analyses and fold  
122 alignment methods with *E. coli* NhaA. This model  
123 has some significant differences from that of  
124 Wakabayashi et al. (Wakabayashi et al. 2000). The  
125 model of Landau suggests that the first two helices  
126 predicted by the Wakabayashi et al. (2000) are  
127 removed and function only as a signal sequence  
128 (Fig. 1). The next 6 TM segments of amino acids  
129 127–315, have the same topology in both models.  
130 Amino acids 328–398, vary in their assignments  
131 between the two models. The last three transmem-  
132 brane segments, TM 10–12 (amino acids 410–500)  
133 are very similar in both models.

134 More recently, Nygaard et al. (Nygaard et al. 2010)  
135 followed up with further structural modeling of  
136 NHE1, based on the structure of NhaA. Their 3D  
137 model principally was similar to the model of  
138 Wakabayashi et al. (Wakabayashi et al. 2000) in  
139 basic transmembrane topology. However, their con-  
140 clusions were not in agreement with some biochemi-  
141 cal data and their EPR results did not distinguish  
142 between the two earlier models, so that the structure  
143 of the NHE1 protein remains unresolved and  
144 controversial (Lee et al. 2011).

- 145 2. *Structural Studies on Human NHE1* – Our labora-  
146 tory has used a “divide and conquer” approach to  
147 study the NHE1 protein (Lee et al. 2011). We stud-  
148 ied individual TM (transmembrane) segments,  
149 which are components of the larger multi-TM  
150 domain. This avoids the difficulties that have

151 plagued researchers in producing multi-TM spanning  
152 proteins. Studies on individual TM helices and  
153 loops have shown that the structures are often very  
154 similar to the structures of helices found in crystal  
155 structures of entire TM proteins (Lee et al. 2011).  
156 We determined the structures of several isolated  
157 TM helices of NHE1 using NMR. These were TM  
158 IV (residues 155–180), TM VI (229–236), TM VII  
159 (250–275), TM IX (338–365), and TM XI  
160 (447–472). Each putative TM segment had unique  
161 structures and none were complete, perfect helices.  
162 Often, discontinuity in the helices was in the middle  
163 of the TM segments and near or containing functionally  
164 important residues. Very briefly, TM IV  
165 was irregularly structured, not resembling  
166 a canonical alpha-helix. Residues 165–168, were  
167 an extended region containing two proline residues.  
168 For TM VI, residues 229–236 contained two helical  
169 regions oriented at approximately right angles to  
170 each other (residues 229–236 and 239–250) surrounding  
171 a central unwound region. TM VII (residues 250–275)  
172 was helical over residues 255–260 and 264–272,  
173 with the central residues at 261–263 non helical. TM IX  
174 (residues 338–365) contained two regions containing  
175 alpha helix structure at residues 340–344 and 353–359,  
176 with a 90° kink at Ser<sup>351</sup> between the two regions which  
177 could provide flexibility. TM XI also contains two  
178 helical regions, residues 447–454 and 460–471 that  
179 are linked by an extended region of residues 455–459  
180 (Lee et al. 2011).  
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## 182 NHE1 Physiological Roles

183 NHE1 has many roles in different cell types (Brett et al.  
184 2005; Fliegel 2008). Knockout of NHE1 from cells  
185 demonstrates a role in growth, which is pronounced  
186 in more acidic media. Similarly, NHE1-knockout mice  
187 demonstrated decreased growth and also ataxia and  
188 epileptic-like seizures. NHE1 is additionally permissive  
189 in cell differentiation and in cell cycle progression.  
190 NHE1 modifies apoptosis. NHE1 is activated by  
191 trophic factor withdrawal in mouse  $\beta$ -cells, leading to  
192 cellular alkalization and progression of apoptosis.  
193 This activation is through p38 dependent phosphorylation  
194 of the NHE1 tail (Malo and Fliegel 2006).

## Pathological Roles

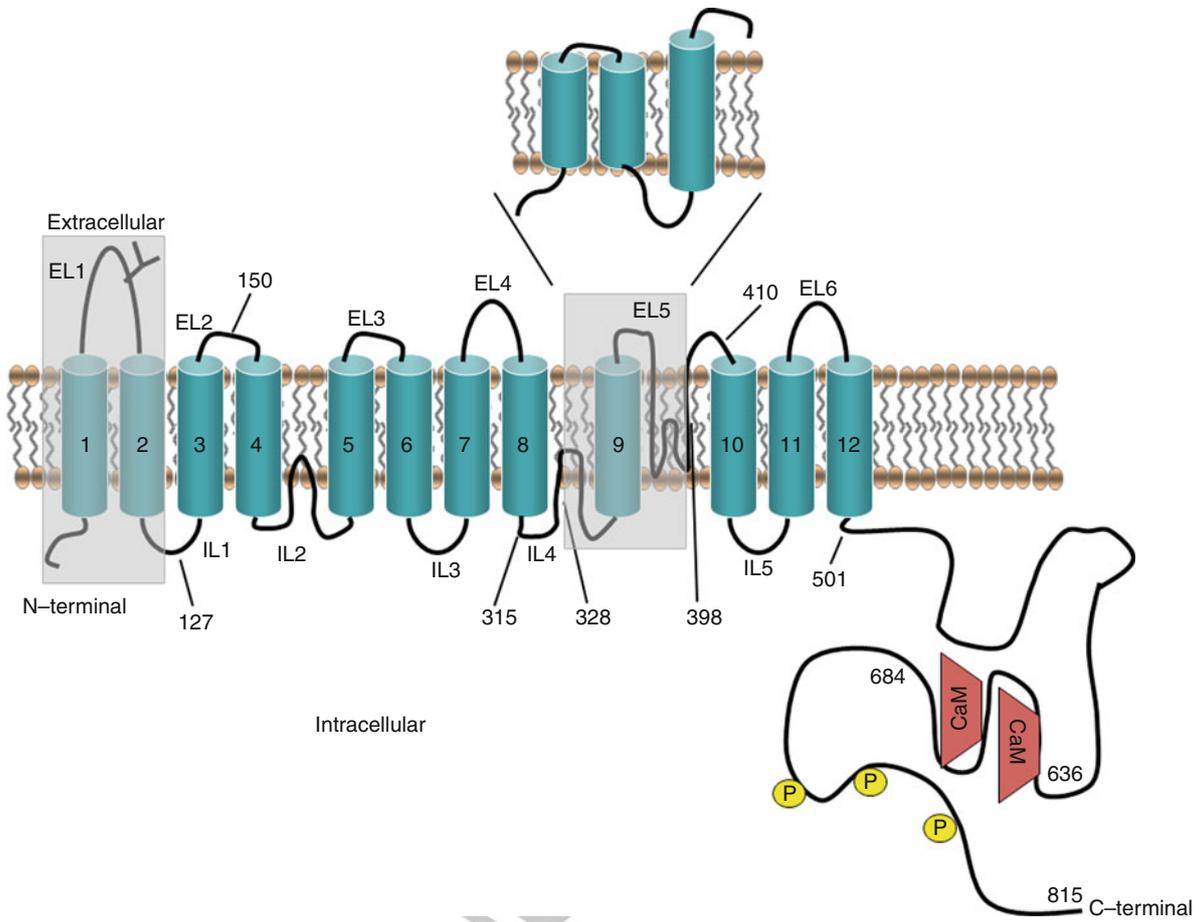
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1. *NHE1 in transformed cells* – The role of NHE1 in  
196 cellular transformation and metastasis is significant.  
197 NHE1-dependent alkalization plays a pivotal role  
198 in the development of a transformed phenotype of  
199 malignant cells, and inhibition of NHE1 prevents or  
200 reduces such development (Stock et al. 2008). This  
201 is particularly well studied in breast cancer. Metas-  
202 tasis is the leading cause of fatality in breast cancer.  
203 In breast cancer cells, NHE1 activation contributes  
204 to cell invasion (Stock et al. 2008). Early work  
205 showed that serum deprivation stimulated NHE1  
206 activity in breast epithelial cell lines, in contrast to  
207 inhibiting it in non-tumor cells. Reshkin and  
208 coworkers championed the idea that a RhoA/  
209 p160ROCK/p38MAPK signaling pathway is  
210 responsible for mediating activation of NHE1  
211 through serum deprivation (Stock et al. 2008).  
212 The mechanism by which NHE1 activity enhances  
213 invasion by breast cancer cells is not only by raising  
214  $\text{pH}_i$ , but also by acidification of the extracellular  
215 microenvironment of tumor cells. The extracellular  
216 acidification is thought to be necessary for protease  
217 activation which facilitates the digestion and  
218 remodeling of the extracellular matrix (Stock et al.  
219 2008), critical in metastasis. 220
2. *NHE1 and heart disease* – NHE is important in  $\text{pH}_i$   
221 regulation in the myocardium. However, it also has  
222 a key role to play in several myocardial pathologies.  
223 The best known of these is the role of NHE1 in  
224 ischemia/reperfusion damage in the myocardium.  
225 During ischemia, anaerobic glycolytic metabolism  
226 is elevated resulting in increased proton production.  
227 This decreases  $\text{pH}_i$  and serves to activate NHE1,  
228 which leads to a rapid accumulation of sodium in  
229 the cell (Fliegel 2008). The high sodium concentra-  
230 tion drives an increase in  $\text{Ca}^{2+}$  via reversal of the  
231 activity of the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger and this triggers  
232 various deleterious pathways leading to cell death.  
233 Many preclinical studies have shown that NHE1  
234 inhibition protects the myocardium from this  
235 calcium overload (Fliegel 2008). Activation of  
236 NHE1 regulatory pathways is also important in  
237 NHE1-mediated damage to the myocardium and  
238 this results in further detrimental activity of the  
239 NHE1 protein (Fliegel 2008). 240
3. *NHE1 in cardiac hypertrophy* – NHE1 is also  
241 important in cardiac hypertrophy (Fliegel 2008). 242

243	Studies have shown that NHE1 inhibition prevents cardiac hypertrophy in several different models of hypertrophy. We also demonstrated that the effect of the hypertrophic agonist aldosterone can be blocked by NHE1 inhibition. A possible mechanism by which NHE1 inhibition prevents hypertrophy is by prevention of increases in intracellular Na <sup>+</sup> . The expression level and activity of NHE1 are elevated in a variety of cardiovascular diseases, including in hypertensive, hypertrophied, or diabetic myocardium (Fliegel 2008) which may accentuate the effects of NHE1 in these diseases.	288
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255	<i>Other cardiovascular conditions</i> – NHE1 inhibitors have been useful in inhibiting diabetic vascular hypertrophy and they also prevent alterations in coronary endothelial function in streptozotocin-induced diabetes. NHE1 inhibitors also have antifibrillatory and antiarrhythmic effects in dogs and rats. Inhibition of NHE1 has also been shown to be protective in cardiac resuscitation models and NHE1 inhibition was beneficial in improving the outcome in a canine transplantation model. NHE1 inhibition may additionally be useful in treatment of circulatory. Overall, NHE1 inhibition has many and varied beneficial cardiovascular effects (Karmazyn et al. 2003).	
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268	<b>Clinical Use of NHE1 Inhibitors</b>	
269	Trials with NHE1 inhibitors have not been very successful. Large-scale studies with various inhibitors have given mostly disappointing results (see (Avkiran et al. 2008) for review). In a small trial of patients with myocardial infarction who received coronary angioplasty, the NHE1 inhibitor cariporide had some beneficial effects on ejection fraction, wall-motion abnormalities and enzyme release. The NHE1 inhibitor eniporide was tested in a larger scale two-stage trial of myocardial infarction. It showed a dose dependent effect to reduce enzyme release, indicating reduced infarction. However, in a second later stage of the trial there was no beneficial effect and an overall negative effect. The GUARDIAN trial tested patients undergoing coronary artery bypass graft surgery. Analysis of subgroups of the trial showed that cariporide was beneficial possibly because treatment with the inhibitor in this trial was early (Avkiran et al. 2008). The EXPEDITION trial tested if the inhibitor	288
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	<b>Summary</b>	
	Na <sup>+</sup> /H <sup>+</sup> exchangers are an important superfamily of cation transporting membrane proteins. There are several families of the monovalent cation proton antiporter (CPA) superfamily. Mammalian NHE1-NHE10 are part of the CPA1 family. They comprise ten distinct isoforms of NHE protein. NHE1 is the best studied. It is a plasma membrane pH regulatory protein that extrudes one intracellular proton in exchange for an extracellular Na <sup>+</sup> . It is comprised of a 500 amino acid membrane domain and a 315 amino acid cytosolic regulatory domain. The topology of NHE1 is in dispute, though the structures of regions of the transmembrane domain have been elucidated. NHE1 is involved heart disease and cancer and is critical in cell growth and differentiation. To date, NHE1 inhibitors have shown great promise in preclinical studies on the myocardium, but these have not translated to beneficial clinical results. Future improvements in the design and use of inhibitors may result in clinically useful treatments.	294
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	<b>Acknowledgments</b> Research by LF in this area is supported by the Canadian Institute of Health Research. LF is supported by an Alberta Ingenuity Medical Scientist award.	313
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	<b>Cross-References</b>	
	▶ Calmodulin	317
	▶ Role of Na Ions in Biological Systems	318
	▶ Sodium Transport in Eucaryotic Cells	319
	▶ Sodium: Physical and Chemical Properties	320
	▶ Types of Na-Binding Sites in Proteins	321
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Galley Proof



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**Na<sup>+</sup>/H<sup>+</sup> Exchangers, Structure, and Function: Role in Human Health and Disease, Fig. 1** Schematic diagram of different versions of the two dimensional topology of NHE1. Alternative two dimensional models of NHE1 topology according to Wakabayashi et al. (2000) and Landau et al. (2000). TM segments according to Wakabayashi et al. (2000) are indicated by 1–12. The carboxyl and amino termini are indicated. Shaded TM's 1–2 are proposed to be deleted by

Landau et al. (2000). The region with varying predicted topology of amino acids 328–398 is indicated by shading and the alternative topology proposed by Landau et al. (2000) is shown above. P indicates putative regulatory phosphorylation sites. CaM indicates regulatory calmodulin binding region. EL and IL refer to intracellular and extracellular loops according to Wakabayashi et al. (2000)

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